



## Assessment of Acid Sulfate Soil Materials (Phase 2) Craignook wetland, South Australia

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Cragnook wetland  
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# CONTENTS

<b>Acknowledgments</b> .....	<b>iv</b>
<b>Executive Summary</b> .....	<b>v</b>
<b>1. Introduction</b> .....	<b>1</b>
<b>2. Laboratory methods</b> .....	<b>5</b>
2.1. Laboratory analysis methods .....	5
2.1.1. Summary of laboratory methods .....	5
2.1.2. Reactive metals method .....	5
2.1.3. Contaminant and metalloid dynamics method .....	5
2.1.4. Monosulfide formation potential method .....	6
2.1.5. Mineral identification by x-ray diffraction .....	6
2.2. Quality assurance and quality control .....	6
<b>3. Results and discussion</b> .....	<b>8</b>
3.1. Summary of soil laboratory results .....	8
3.1.1. Reactive metals data .....	8
3.1.2. Contaminant and metalloid dynamics data .....	8
3.1.3. Monosulfide formation potential data .....	14
3.1.4. Mineral identification by x-ray diffraction .....	16
3.2. Interpretation and discussion of results .....	16
<b>4. Risk assessment</b> .....	<b>19</b>
4.1. Risk assessment framework .....	19
4.2. Assessment of risks .....	21
4.2.1. Risks associated with acidification .....	21
4.2.2. Risks associated with contaminant mobilisation .....	22
4.2.3. Risks associated with de-oxygenation .....	22
<b>5. Broad acid sulfate soil management options</b> .....	<b>24</b>
<b>References</b> .....	<b>26</b>
<b>Appendices</b> .....	<b>28</b>
<b>Appendix 1 Reactive metals data</b> .....	<b>29</b>
<b>Appendix 2 Contaminant and metalloid dynamics data</b> .....	<b>30</b>
<b>Appendix 3 Monosulfide formation potential data</b> .....	<b>31</b>

## LIST OF FIGURES

Figure 1-1 Craignook wetland aerial photograph with Phase 1 sampling sites identified.....	3
Figure 3-1 Contaminant and metalloid dynamics results for Craignook wetland soil materials for pH, SEC, Eh, silver (Ag), aluminium (Al) and arsenic (As).....	11
Figure 3-2 Contaminant and metalloid dynamics results for Craignook wetland soil materials for cadmium (Cd), cobalt (Co), chromium (Cr), copper (Cu), iron (Fe) and manganese (Mn). .....	12
Figure 3-3 Contaminant and metalloid dynamics results for Craignook wetland soil materials for nickel (Ni), lead (Pb), antimony (Sb), selenium (Se), vanadium (V) and zinc (Zn). ....	13
Figure 3-4 Selected trace elements plotted against pH.....	14
Figure 3-5 pH and Eh dynamics during monosulfide formation potential tests in surface soil sample CNK 1.1 from Craignook wetland.....	15

## LIST OF TABLES

Table 1-1 Priority ranking criteria adopted by the Scientific Reference Panel of the Murray-Darling Basin Acid Sulfate Soils Risk Assessment Project, from MDBA (2010).....	2
Table 1-2 Rationale for Phase 2 sample selection, from MDBA (2010). ....	4
Table 1-3 Summary of Craignook wetland samples analysed for Phase 2 assessment. ....	4
Table 2-1 Phase 2 data requirements - list of parameters and objective for conducting the test, from MDBA (2010). ....	5
Table 3-1 Craignook reactive metals data. ....	8
Table 3-2 Summary of contaminant and metalloid dynamics data ....	9
Table 3-3 Summary of monosulfide formation potential data for the Craignook wetland surface soil material CNK 1.1 after 7 weeks (3.6 g/L sucrose). ....	15
Table 3-4 Summary of the degree of hazard associated with the measured contaminant and metalloid concentrations in the Craignook wetland. ....	17
Table 3-5 Guideline thresholds for the degree of hazard associated with acid volatile sulfide (S <sub>AV</sub> ) concentrations. ....	18
Table 4-1 Standardised table used to determine the consequences of a hazard occurring, from MDBA (2011). ....	19
Table 4-2 Likelihood ratings for the disturbance scenario, from MDBA (2011). ....	20
Table 4-3 Risk assessment matrix, adapted from Standards Australia & Standards New Zealand (2004). ....	20
Table 4-4 Summary of risks associated with acid sulfate soil materials in the Craignook wetland. ....	23
Table 5-1 Summary of management options and possible activities, from EPHC & NRMCC (2011). ....	25

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## EXECUTIVE SUMMARY

An initial Phase 1 acid sulfate soil investigation of the Craignook wetland during September 2008 showed acid sulfate soils to be a priority concern within this wetland complex. Based on Phase 1 recommendations, a Phase 2 investigation was undertaken for the Craignook wetland to determine the nature, severity and the specific risks associated with acid sulfate soil materials.

The 24 hour **reactive metals** tests were undertaken to determine those metals and metalloids extractable with a moderately strong acid i.e. potentially available from binding sites on soil minerals such as iron (Fe), manganese (Mn) and aluminium (Al) oxides. Although comparisons can be made with soil and sediment quality guidelines, these are defined for total concentrations and not partial extractions. The results showed that concentrations were below the sediment quality guidelines (SQG) and soil ecological investigation levels (EIL) for those elements where guidelines are available. The concentrations for this reactive metals partial extraction are considered to be moderately high for iron (Fe).

The **contaminant and metalloid dynamics** (CMD) tests were undertaken to assess the release of metals during a water extraction, and to assess dynamics in response to saturation over time by incubating soil materials for periods of 1, 7, 14 and 35 days. The degree to which metal and metalloid concentrations exceed ANZECC/ARMCANZ environmental protection guideline values for environmental protection was used to characterise the degree of hazard. The soils were mainly cracking clay soils with marginal sands. The soils remained moderately acidic for the most part, but the surface of the clay soil showed a more significant increase and a larger decrease in Eh to reducing conditions. The highest concentrations of metals and metalloids were found in this more reducing soil. For Craignook wetland, cobalt (Co) and iron (Fe) were assigned a moderate hazard with concentrations exceeding ANZECC/ARMCANZ environmental protection guidelines by more than 10 times. Aluminium (Al), arsenic (As), nickel (Ni) and zinc (Zn) were also significantly above ANZECC/ARMCANZ environmental protection guideline values and represent significant hazards. The dominant control on metal solubility during the contaminant and metalloid dynamics tests was Eh. All samples showed a decrease in Eh, but only one sample decreased sufficiently for iron (Fe) to become soluble. The increase in iron (Fe) was associated with the release of both metals and metalloids e.g. arsenic (As), probably due to the reductive dissolution of iron oxides/oxyhydroxides. Iron (Fe) and arsenic (As) were still increasing at the end of the 35 day contaminant and metalloid dynamics experiments.

The Craignook wetland has been classified as medium conservation status by the SA Murray–Darling Basin Natural Resources Management Board (Miles *et al.* 2010). The main hazards considered in this study that may impact on wetland values are acidification, contaminant mobilisation and deoxygenation. The wetland has been allocated a **medium** risk rating for **acidification** and a **high contaminant mobilisation** risk rating for **soils**. For **surface waters**, the risk is largely dependent on surface and sub-surface hydrology and is thus scenario dependent. Taking into account the range of likely scenarios, from very low flows (highest risk) to very high flows (lowest risk), the risk to surface waters in the wetland has been allocated a **low to high** risk rating for **acidification** and a **medium to high** risk rating for **contaminant mobilisation**. The risk associated with **deoxygenation** is considered **high** due to the detection of monosulfides in one sample and very high water soluble sulfate concentrations in most samples. Very high dissolved sulfide levels were also present, and precipitation of monosulfides or possibly pyrite had occurred by the end of the 7 week monosulfide formation potential incubation.

In designing a management strategy for dealing with acid sulfate soils in Craignook wetland, other values and uses of the wetland need to be taken into account to ensure that any intervention is compatible with other management plans and objectives for the wetland.

The wetland soils in places were dry but pools of water had developed in small areas. Previous drying had led to the development of extensive cracking of the soils. Management options considered should relate to controlling or treating acidification and the protection of connected or adjacent wetlands. Due to the medium to high risks to the wetland values associated with acidification, contaminant mobilisation and deoxygenation in Craignook wetland, a monitoring program is strongly recommended.



# 1. INTRODUCTION

At its March 2008 meeting, the Murray–Darling Basin Ministerial Council discussed the emerging issue of inland acid sulfate soils and the associated risks to Murray–Darling Basin waterways and agreed that the extent of the threat posed by this issue required assessment. The purpose of the Murray–Darling Basin Acid Sulfate Soils Risk Assessment Project was to determine the spatial occurrence of, and risk posed by, acid sulfate soils at priority wetlands in the River Murray system, wetlands listed under the Ramsar Convention on Wetlands of International Importance and other key environmental sites in the Murray–Darling Basin. The project involved the selection of wetlands of environmental significance, as well as those that may pose a risk to surrounding waters. These wetlands were then subjected to a tiered assessment program, whereby wetlands were screened through a desktop assessment stage, followed by a rapid on-ground appraisal, and then detailed on-ground assessment if results of previous stages indicated an increased likelihood of occurrence of acid sulfate soils.

Detailed assessments of acid sulfate soils within the Murray-Darling Basin (MDB) are conducted as a two-phase process under the MDB Acid Sulfate Soil Risk Assessment Project (ASSRAP). Phase 1 investigations are initially undertaken to determine whether or not acid sulfate soil materials are present in the study area, and provide characterisation of the properties and types of acid sulfate soils. Phase 2 investigations are only conducted if the acid sulfate soil materials from Phase 1 are determined to be a priority concern for the study area and, based on Phase 1 recommendations, selected samples undergo further investigations to determine the nature, severity and the specific risks associated with the acid sulfate soil materials. Phase 2 activities include: (i) soil laboratory analysis to confirm and refine the hazards associated with contaminant mobilisation and/or deoxygenation, (ii) a risk assessment, and (iii) interpretation and reporting, including discussion on broad acid sulfate soil management options.

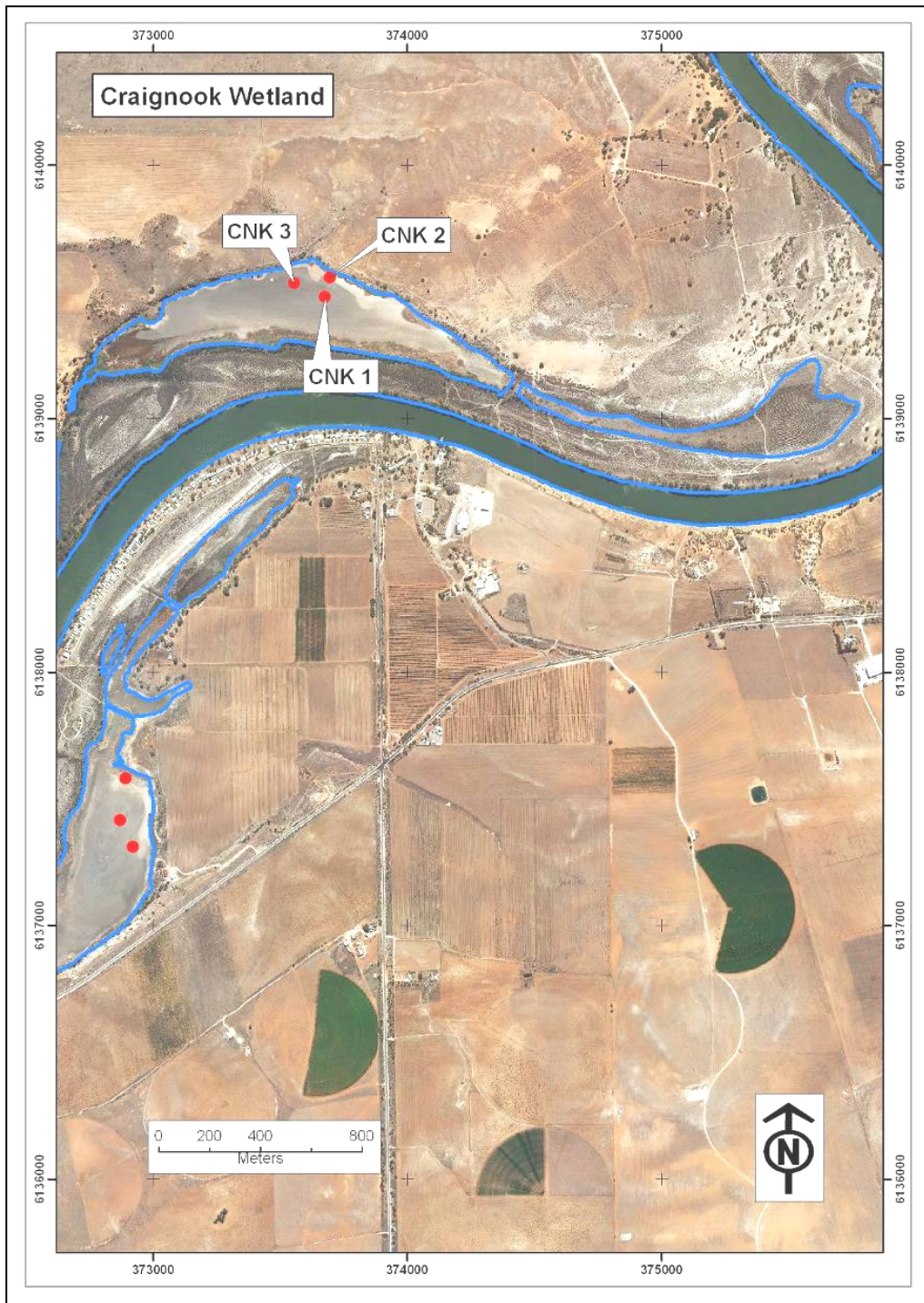
Detailed Phase 1 acid sulfate soil assessments were undertaken at almost 200 wetlands and river channels throughout the Murray-Darling Basin. In South Australia, 56 wetlands along the River Murray between Lock 1 and Lock 5 were investigated by CSIRO Land and Water (Grealish *et al.* 2010). From these Phase 1 investigations, 13 wetlands were selected for further investigation. Nearly all of the wetlands along the River Murray between Wellington and Blanchetown (Lock 1) in South Australia also received detailed Phase 1 acid sulfate soil assessments (Grealish *et al.* 2011) and of these 23 wetlands were selected for further investigation in Phase 2. This included some wetlands below Lock 1 from earlier studies (Fitzpatrick *et al.* 2008; Fitzpatrick *et al.* 2010).

Following the Craignook wetland Phase 1 assessment (Grealish *et al.* 2011) and the priority ranking criteria adopted by the Scientific Reference Panel of the MDB ASSRAP (see Table 1-1), Craignook wetland was selected for Phase 2 detailed assessment.

The Phase 1 assessment sampled from 3 sites (Figure 1-1) from the wetland which comprised cracking clay soils and marginal sands. The wetland was partially dry at the time of sampling. Sites CNK 1-3 were sampled from the central part of the wetland. The Phase 1 assessment identified 1 high priority site based on the presence of sulfuric materials, 2 high priority sites based on the presence of hypersulfidic materials, 2 high priority sites based on hyposulfidic materials with  $S_{CR} \geq 0.10\%$  and 2 moderate priority sites based on the presence of hyposulfidic materials with  $S_{CR} < 0.10\%$ . Phase 2 investigations were carried out on selected surface soil samples from two sites (CNK 1 and CNK 2) identified in the Phase 1 assessment (Grealish *et al.* 2011).

**Table 1-1 Priority ranking criteria adopted by the Scientific Reference Panel of the Murray-Darling Basin Acid Sulfate Soils Risk Assessment Project, from MDBA (2010).**

Priority	Soil material
<b>High Priority</b>	<p>All sulfuric materials.</p> <p>All hypersulfidic materials (as recognised by either 1) incubation of sulfidic materials or 2) a positive net acidity result with a Fineness Factor of 1.5 being used).</p> <p>All hyposulfidic materials with <math>S_{CR}</math> contents <math>\geq 0.10\%</math> S.</p> <p>All surface soil materials (i.e. within 0-20 cm) with water soluble sulfate (1:5 soil:water) contents <math>\geq 100 \text{ mg kg}^{-1} \text{ SO}_4</math>.</p> <p>All monosulfidic materials.</p>
<b>Moderate Priority</b>	<p>All hyposulfidic materials with <math>S_{CR}</math> contents <math>&lt; 0.10\%</math> S.</p>
<b>No Further Assessment</b>	<p>Other acidic soil materials.</p> <p>All other soil materials.</p>



**Figure 1-1 Craignook wetland aerial photograph with Phase 1 sampling sites identified.**

A summary of the soil laboratory analyses undertaken as part of the Phase 2 assessment and the sample selection criteria for each analysis is given in Table 1-2. Soil samples identified to undergo Phase 2 laboratory analysis are primarily from the surface and near-surface layers, as these are the soils most likely to have initial contact with water. A list of the samples selected for Phase 2 analysis for the Craignook wetland is presented in Table 1-3.

**Table 1-2 Rationale for Phase 2 sample selection, from MDBA (2010).**

Parameter	Samples selected
Reactive metals	Conducted on selected upper two surface samples.
Contaminant and metalloid dynamics	Conducted on selected upper two surface samples.
Monosulfide formation potential	Conducted on surface samples of dry sites that meet the water extractable sulfate criteria for monosulfides.
Mineral identification by X-ray diffraction (XRD)	Conducted on a limited number of selected crystals and minerals (if present). Most likely to be associated with sulfuric layers to confirm acid mineral presences.
Acid base accounting data	Conducted only on samples from wetlands below Lock 1 and Burnt Creek/Loddon River if not previously analysed and $pH_{KCl} < 4.5$ .

**Table 1-3 Summary of Craignook wetland samples analysed for Phase 2 assessment.**

Soil Laboratory Test	Craignook wetland samples	Sample depth (cm)	Number of samples analysed
Reactive metals	CNK1.1	0-10	4
	CNK1.2	10-20	
	CNK2.1	0-5	
	CNK2.2	5-35	
Contaminant and metalloid dynamics	CNK1.1	0-10	4
	CNK1.2	10-20	
	CNK2.1	0-5	
	CNK2.2	5-35	
Monosulfide formation potential	CNK1.1	0-10	1
Mineral identification by X-ray diffraction (XRD)	-		0

## 2. LABORATORY METHODS

### 2.1. Laboratory analysis methods

#### 2.1.1. Summary of laboratory methods

A list of the method objectives for the Phase 2 assessment are summarised below in Table 2-1. All soil samples analysed in this Phase 2 assessment were collected and subsequently stored as part of the Phase 1 field assessment.

**Table 2-1 Phase 2 data requirements - list of parameters and objective for conducting the test, from MDBA (2010).**

Parameter	Objective
Reactive metals	Assists with determining impacts on water quality by determining weakly to moderately strongly bound metals.
Contaminant and metalloid dynamics	Assists with determining impacts on water quality by simulating longer time frames that create anaerobic conditions. Identifies metal release concentrations that may occur over a 5 week time frame.
Monosulfide formation potential	Determine relative propensity for monosulfides to form following inundation.
Mineral identification by X-ray diffraction (XRD)	Characterisation and confirmation of minerals present.

Guidelines on the approaches that were followed as part of this Phase 2 assessment are presented in full in the detailed assessment protocols (MDBA 2010).

#### 2.1.2. Reactive metals method

The guidelines for the reactive metals method are outlined as an addendum to the detailed assessment protocols (MDBA 2010). In this method, samples were prepared by disaggregation (not grinding) using a jaw crusher, and then sieved to include only the <2 mm fine earth fraction. A total of 2.5 g soil was added to 40 ml of 0.1 M HCl, gently mixed for 1 hour and filtered through a pre-acid washed 0.45 µm nitro-cellulose filter. The metals examined comprised silver (Ag), aluminium (Al), arsenic (As), cadmium (Cd), cobalt (Co), chromium (Cr), copper (Cu), iron (Fe), manganese (Mn), nickel (Ni), lead (Pb), antimony (Sb), selenium (Se), vanadium (V) and zinc (Zn).

#### 2.1.3. Contaminant and metalloid dynamics method

The guidelines for the contaminant and metalloid dynamics method are outlined in Appendix 7 of the detailed assessment protocols (MDBA 2010). The contaminant and metalloid dynamics method was designed to determine the release of metals and metalloids in soils after 24 hours. The data represent the availability of metals and metalloids from a weak extraction (water, and thus easily bioavailable) of saturated soils, and for dry wetland soils, those easily mobilised from mineral surfaces and readily soluble mineral phases (such as salts). The exercise was repeated in a batch process for longer time periods (7 days, 14 days and 35 days). The latter approach was aimed at understanding changes in concentrations over time. This is particularly important for dried soils which have been in contact with the

atmosphere. The soil materials and the release/uptake of metals/metalloids are expected to change as the chemical environment changes from oxidising to reducing. The data can be compared to existing water quality guidelines, although care should be taken when extrapolating to surface waters without knowledge of hydrological conditions and natural chemical barriers. The impact on surface waters will be governed by the upward chemical flux which is a function of soil type, water flow, diffusion and the chemistry of the soils near the sediment-water interface.

Redox potential (Eh) and pH were determined using calibrated electrodes linked to a TPS WP-80 meter; Eh measurements were undertaken in an anaerobic chamber to minimise the rapid changes encountered due to contact with the atmosphere, and are presented relative to the standard hydrogen electrode (SHE). Specific electrical conductance (SEC) was determined using a calibrated electrode linked to a TPS WP-81 meter. All parameters were measured on filtered (0.45 µm) water samples.

#### **2.1.4. Monosulfide formation potential method**

The guidelines for the monosulfide formation potential method are outlined in Appendix 8 of the detailed assessment protocols (MDBA 2010). In this study 3.6 g/L sucrose was used as an organic substrate instead of the 7.2 g/L outlined in the protocols. In addition to sampling after seven weeks, water samples were collected and analysed immediately after inundating the soils (i.e. Day 0). The pore-water pH and Eh were determined at Day 0.

The reactive iron (Fe) fraction in field moist sediments was extracted using 1.0 M HCl (Claff *et al.* 2010). The ferrous iron ( $\text{Fe}^{2+}$ ) and total iron ( $\text{Fe}^{2+} + \text{Fe}^{3+}$ ) fractions were immediately fixed following extraction. The ferrous iron trap was made up from a phenanthroline solution with an ammonium acetate buffer (APHA 2005), and the total iron trap also included a hydroxylamine solution (APHA 2005). The iron species were quantified colorimetrically using a Hach DR 2800 spectrophotometer.

Redox potential and pH were determined using calibrated electrodes linked to a TPS WP-80 meter; Eh measurements are presented versus the standard hydrogen electrode. In this study the solid phase elemental sulfur fraction was extracted using toluene as a solvent and quantified by high-performance liquid chromatography (HPLC) (McGuire and Hamers 2000). Pore-water sulfide was preserved in zinc acetate prior to determination by the spectrophotometric method of Cline (1969).

#### **2.1.5. Mineral identification by x-ray diffraction**

The guidelines for mineral identification by x-ray diffraction are outlined in the detailed assessment protocols (MDBA 2010).

## **2.2. Quality assurance and quality control**

For all tests and analyses, the quality assurance and quality control procedures were equivalent to those endorsed by NATA (National Association of Testing Authorities). The standard procedures included the monitoring of blanks, duplicate analysis of at least 1 in 10 samples, and the inclusion of standards in each batch.

Reagent blanks and method blanks were prepared and analysed for each method. All blanks examined here were either at, or very close to, the limits of detection. On average, the frequencies of quality control samples processed were: 10% blanks, 10% laboratory duplicates, and 10% laboratory controls. The analytical precision was  $\pm 10\%$  for all analyses. In addition, for all samples, reactive metals and contaminant and metalloid dynamics tests were duplicated. For the reactive metals, two International Standards (Reference Stream Sediment STSD-2 and STSD-3 Canadian Certified Reference Materials) were processed in

an identical manner to the samples. Precision was excellent with the coefficient of variation (standard deviation/mean\*100) typically being in the range < 1 to 2 %.

### 3. RESULTS AND DISCUSSION

#### 3.1. Summary of soil laboratory results

##### 3.1.1. Reactive metals data

The data are presented on a dry weight basis (mg kg<sup>-1</sup>) and shown in Table 3-1. The 24 hour reactive metals studies provide an indication of those metals and metalloids which are more strongly bound to minerals (or weakly soluble with an acid extraction) than would be soluble with a water extraction, and thus have the potential to be released. The use of a moderately strong acid (0.1 M HCl) should provide an indication of “stored metals” and metalloids associated with iron (Fe) and manganese (Mn) oxides and organic materials as well as acid soluble minerals. It is commonly found that the concentrations of metals and metalloids released using extractions are much higher than those found in solution (Goody *et al.* 1995). Although guideline values exist for soils and sediments, these are generally for total soil concentrations, and therefore, are not directly appropriate for the data from metal mobilisation studies. Nevertheless, they provide a basis for comparison; and concentrations close to or above guideline values indicate an elevated hazard.

The concentrations of metals and metalloids were below sediment quality guideline (SQG) values and soil ecological investigation levels (EIL) for those elements for which guidelines are available. The concentrations for most reactive metals and metalloids are relatively low, but relatively high for iron (Fe) (Table 3-1).

**Table 3-1 Craignook reactive metals data.**

Concentrations in mg kg<sup>-1</sup>, and µg kg<sup>-1</sup> as indicated by asterisk.

Sample	Ag*	Al	As	Cd*	Co	Cr*	Cu	Fe	Mn	Ni	Pb	Sb*	Se*	V	Zn
CNK 1.1	6.5	372	2.1	98	3.1	118	4.9	1006	255	5.9	3.8	< 2.0	58	12	8.3
CNK 1.2	6.2	298	1.8	49	0.58	76	3.6	983	41	3.9	2.5	2.4	39	10	3.7
CNK 2.1	0.34	42	0.60	1.0	0.21	21	0.25	78	3.1	0.41	0.04	< 1.7	2.8	0.25	0.31
CNK 2.2	1.2	50	0.46	2.2	0.35	29	0.31	137	4.1	0.47	0.15	< 1.8	4.4	0.31	0.36
<sup>1</sup> SQG	1000	-	20	1500	-	80000	65	-	-	21	50	2000	-	-	200
<sup>2</sup> Soil EIL	-	-	20	3000	-	-	100	-	500	60	600	-	-	50	200

\* Units are in µg kg<sup>-1</sup>

< value is below detection limit

<sup>1</sup>SQG: Sediment Quality Guideline Value (Australian and New Zealand Guidelines for Fresh and Marine Water Quality 2000)

<sup>2</sup>Soil EIL: Soil – Ecological Investigation Level (Australian and New Zealand Guidelines for Fresh and Marine Water Quality 2000)

##### 3.1.2. Contaminant and metalloid dynamics data

The contaminant and metalloid dynamics data for the four Craignook wetland soil materials examined are presented in Appendix 2, summarised in Table 3-2 and plotted against time in Figure 3-1 to Figure 3-3. Table 3-2 also compares the pore-water metal contents to the relevant national water quality guideline for environmental protection (ANZECC/ARMCANZ 2000).

The contaminant and metalloid dynamics data for the Craignook wetland soil materials are summarised in Table 3-2. The table also compares the pore-water concentrations to the relevant national water quality guideline for environmental protection (ANZECC/ARMCANZ 2000).



**Table 3-2 Summary of contaminant and metalloid dynamics data**

Parameter	units	ANZECC Guidelines	Craignook		
			Min.	Median	Max.
pH		6.5-8.0	4.4	5.3	6.5
EC*	$\mu\text{S cm}^{-1}$	2200	35	398	3650
Eh	mV	-	79	427	547
Ag	$\mu\text{g l}^{-1}$	0.05	<0.01	<0.01	<0.2
Al <sup>A</sup>	$\text{mg l}^{-1}$	0.055	<0.05	<0.05	0.14
As <sup>B</sup>	$\mu\text{g l}^{-1}$	13	<0.3	0.37	45
Cd	$\mu\text{g l}^{-1}$	0.2	<0.01	<0.1	0.50
Co	$\mu\text{g l}^{-1}$	2.8	1.8	3.0	29
Cr <sup>C</sup>	$\mu\text{g l}^{-1}$	1	<0.07	<0.3	<3
Cu <sup>H</sup>	$\mu\text{g l}^{-1}$	1.4	<0.20	<1	<10
Fe <sup>I</sup>	$\text{mg l}^{-1}$	0.3	<0.10	<0.10	23
Mn	$\mu\text{g l}^{-1}$	1700	28	174	6672
Ni <sup>H</sup>	$\mu\text{g l}^{-1}$	11	1.3	4.8	20
Pb <sup>H</sup>	$\mu\text{g l}^{-1}$	3.4	<0.20	<3	<6
Sb	$\mu\text{g l}^{-1}$	9	<0.60	<2	<20
Se	$\mu\text{g l}^{-1}$	11	<0.03	<0.08	1.0
V	$\mu\text{g l}^{-1}$	6	<0.08	0.30	4.1
Zn <sup>H</sup>	$\mu\text{g l}^{-1}$	8	<1	2.1	19

**Exceeded  
ANZECC  
Guideline (x1)**

**Exceeded  
ANZECC  
Guideline (x10)**

**Exceeded  
ANZECC  
Guideline (x100)**

**Notes.**

The ANZECC guideline values for toxicants refer to the Ecosystem Protection – Freshwater Guideline for protection of 95% of biota in ‘slightly-moderately disturbed’ systems, as outlined in the Australian Water Quality Guidelines for Fresh and Marine Water Quality (ANZECC/ARMCANZ 2000).

\* ANZECC water quality upper guideline ( $125\text{-}2200 \mu\text{S cm}^{-1}$ ) for freshwater lowland rivers in South-east Australia is provided for salinity (there are currently no trigger values defined for ‘Wetlands’.)

<sup>A</sup> Guideline is for Aluminium in freshwater where pH > 6.5.

<sup>B</sup> Guideline assumes As in solution as Arsenic (AsV).

<sup>C</sup> Guideline for Chromium is applicable to Chromium (CrVI) only.

<sup>H</sup> Hardness affected (refer to Guidelines).

<sup>I</sup> Fe Guideline for recreational purposes.

The pH of the samples were initially moderately acidic, and although most samples increased slightly in pH, three of the profiles remained acidic with pH of ca. pH 5.3 (Figure 3-1). The other sample increased to pH 6.5, the lower limit of the ANZECC/ARMCANZ environmental protection guideline value. The SEC showed a wide range, with samples from profile CNK 2 (sands on the margin of the wetland) being much fresher than the clay soil CNK 1. The SEC showed no consistent trend (Figure 3-1).

The Eh of the soils decreased with time for all samples. In one surface sample (CNK 1.1), the Eh became moderately reducing, whilst in the others it remained moderately oxidising.

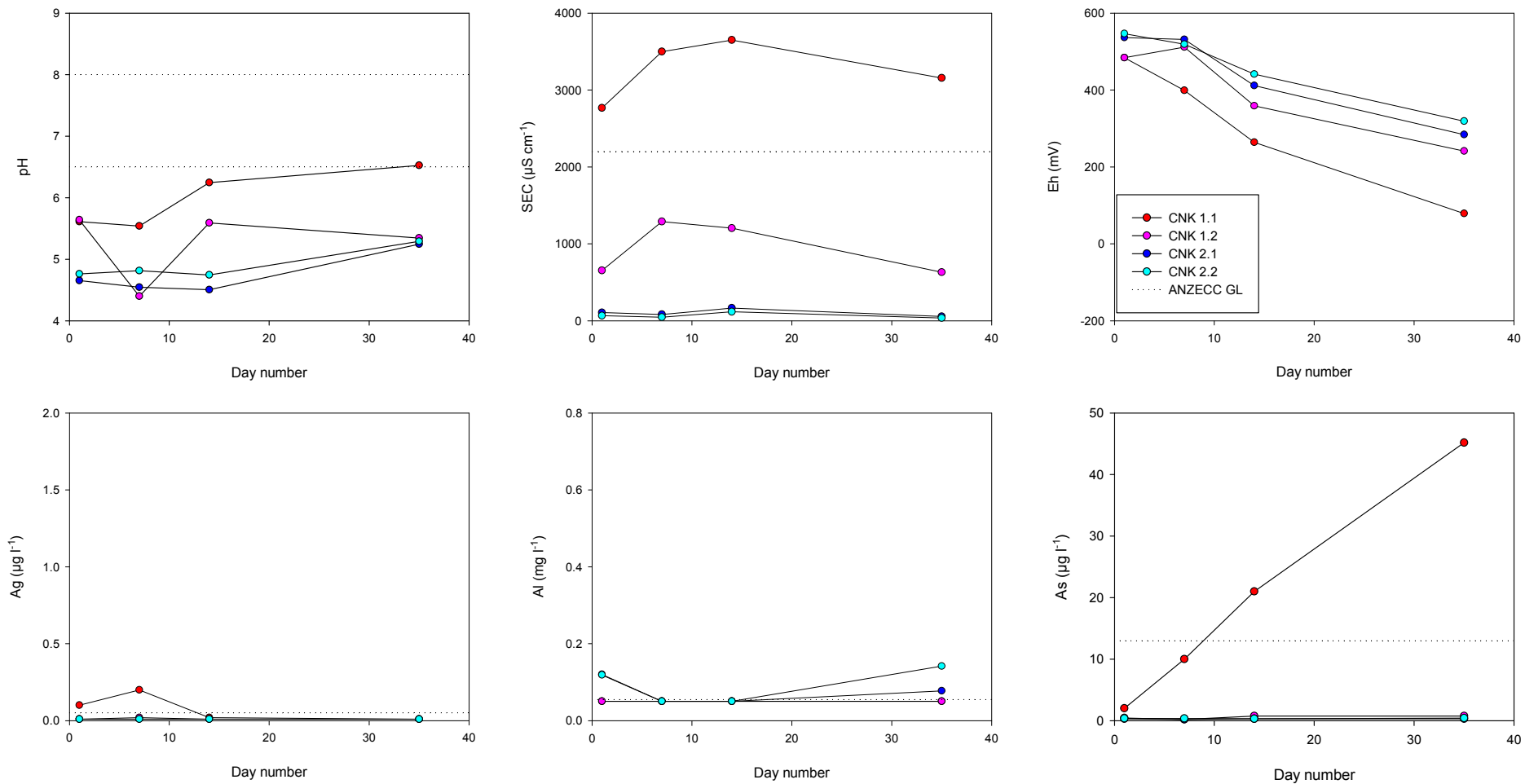
Iron (Fe) concentrations were initially low after day 1, and remained so in the more oxidising samples. In the reducing sample, iron increased consistently, reaching very high concentration ( $23 \text{ mg l}^{-1}$ ) by day 35. Manganese (Mn) was also high in the reducing sample, and both iron and manganese were higher than the ANZECC/ARMCANZ environmental protection guideline value.

Aluminium concentrations were generally low, but breached the ANZECC/ARMCANZ environmental protection guideline value high in samples from the sandy soil profile CNK 2 (Figure 3-1). Arsenic (As) concentrations increased significantly in sample CNK 1.1, and showed a trend similar to iron. It breached the ANZECC/ARMCANZ environmental protection guideline value on day 14, and continued to increase until at day 35 it reached  $45 \mu\text{g l}^{-1}$ . Vanadium (V) remained below the ANZECC/ARMCANZ environmental protection guideline value in all samples (Figure 3-3).

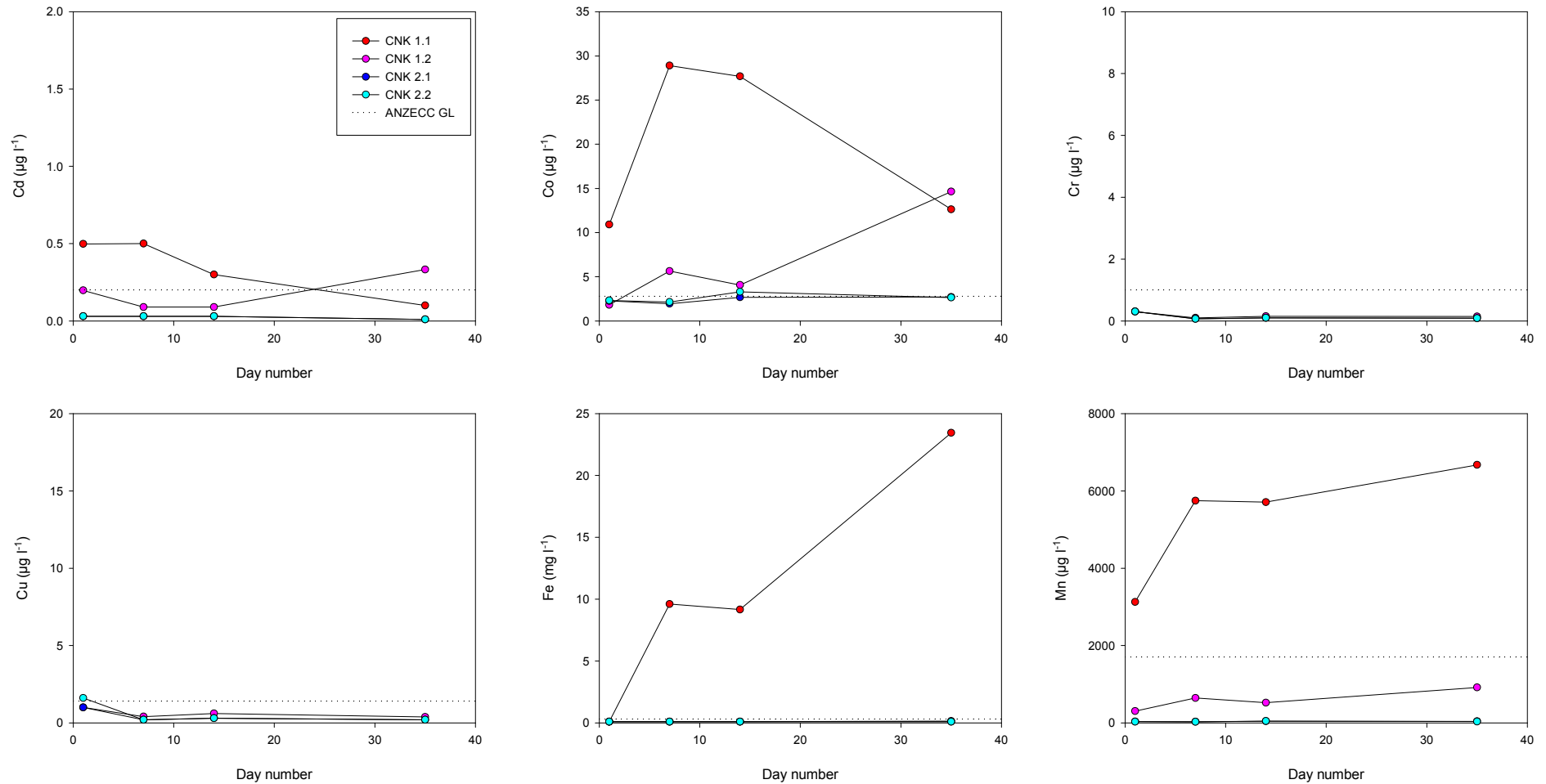
For the metals, the highest concentrations were typically present in soil sample from the clayey profile CNK 1, with elevated cobalt (Co), nickel (Ni) and zinc (Zn), and to a lesser degree cadmium (Cd) (Figure 3-1 to Figure 3-3).

The magnitude of metal mobilisation is affected by many factors that include but are not exclusive to: 1) the abundance and form of metal and metalloid contaminants; 2) the abundance and lability of organic matter; 3) the abundance and reactivity of iron minerals; 4) availability of sulfate; 5) acid/alkalinity buffering capacity; 6) pH; 7) Eh; 8) clay content; 9) microbial activity; 10) temperature; and 11) porosity (MDBA 2010).

Most metals display a trend with pH (Figure 3-4), with most metals (except aluminium) being higher at pH above pH 5.5. The elevated concentrations of arsenic (As), cobalt (Co), nickel (Ni), vanadium (V) and zinc (Zn) are similar to iron (Fe) and manganese (Mn) suggesting a control from the reductive dissolution of oxides/oxyhydroxides of these metals.

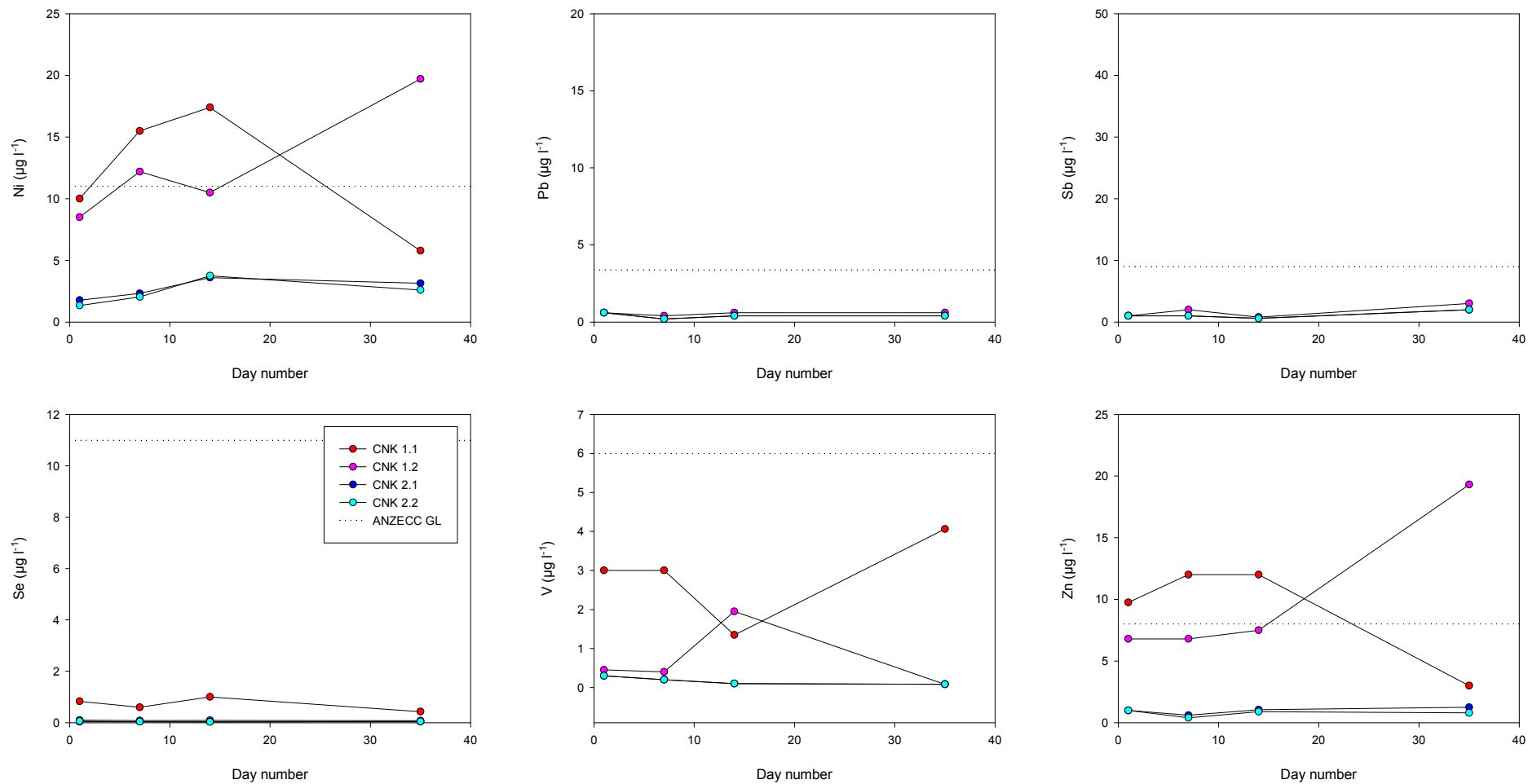


**Figure 3-1 Contaminant and metalloid dynamics results for Cragnook wetland soil materials for pH, SEC, Eh, silver (Ag), aluminium (Al) and arsenic (As).**  
 Note: silver (Ag) was all < detection limit, data represent detection limits which vary according to required dilutions.



**Figure 3-2 Contaminant and metalloid dynamics results for Caignook wetland soil materials for cadmium (Cd), cobalt (Co), chromium (Cr), copper (Cu), iron (Fe) and manganese (Mn).**

Note: chromium (Cr) and copper (Cu) were all < detection limit, data represent detection limits which vary according to required dilutions.



**Figure 3-3 Contaminant and metalloid dynamics results for Cragnook wetland soil materials for nickel (Ni), lead (Pb), antimony (Sb), selenium (Se), vanadium (V) and zinc (Zn).**

Note: lead (Pb) and antimony (Sb) were all < detection limit, data represent detection limits which vary according to required dilutions.

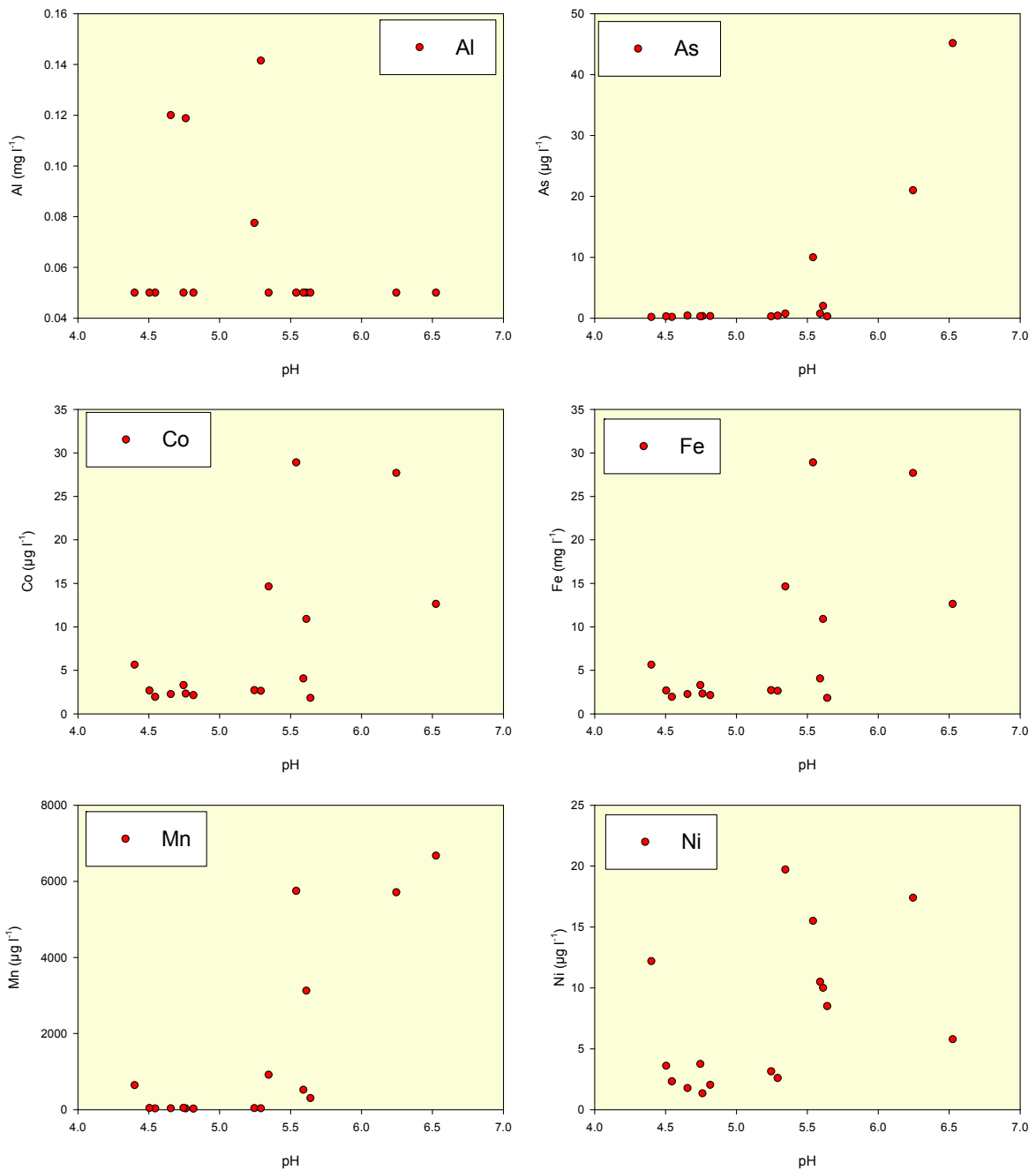


Figure 3-4 Selected trace elements plotted against pH.

### 3.1.3. Monosulfide formation potential data

The monosulfide formation potential data for sample CNK 1.1 are shown in Table 3-3. The pH of the soil water changed from 5.48 to 4.71 over the seven week incubation period (Figure 3-5).

The decrease in pH is different from the contaminant and metalloid dynamics tests, where pH increased from 5.61 to 6.53 (Figure 3-1). This may be due to fermentation of organic substrate added (sucrose) which caused acidification of the pore-waters.

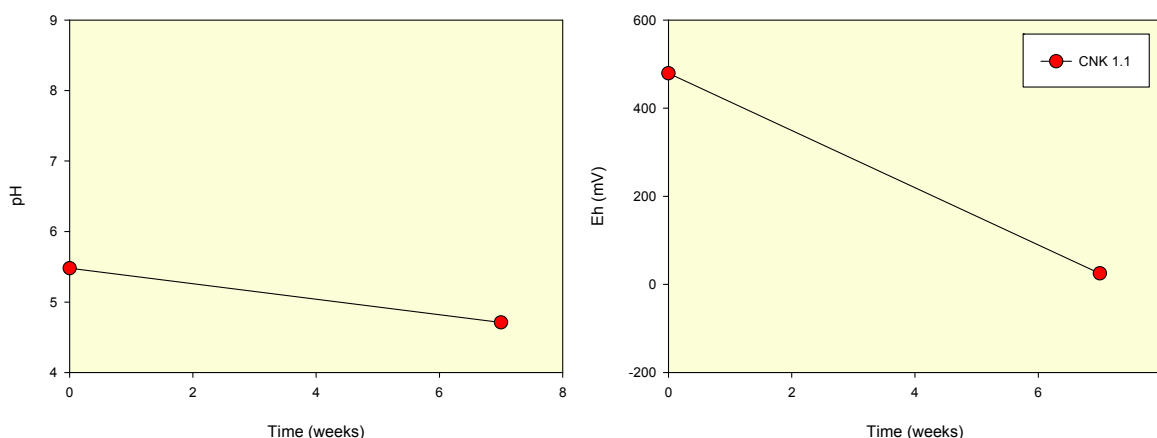
The Eh decreased significantly from 479 to 25 mV (Table 3-3) indicating a change to more reducing conditions. The Eh data are similar to the contaminant and metalloid dynamics experiments where Eh decreased from 484 to 79 mV.

**Table 3-3 Summary of monosulfide formation potential data for the Craignook wetland surface soil material CNK 1.1 after 7 weeks (3.6 g/L sucrose).**

Inundation Time	Parameter	Units	Craignook (CNK 1.1)
Day 0	Total Fe	mg/kg	5978
	Fe(II) <sup>-</sup>	mg/kg	1268
	Sulfate*	mg/kg	5905
	pH		5.48
	Eh	mV	479
Week 7	pH		4.71
	Eh	mV	25
	S <sub>AV</sub>	Wt. %S	0.01
	S <sup>0</sup>	Wt. %S	0.07
	Pyrite-S	Wt. %S	0.10
	Dissolved S <sup>2-</sup>	µg/L	50189

\* completed during Phase 1

After 7 weeks, acid volatile sulfide (S<sub>AV</sub>) was present, but at very low concentrations (0.01 %), and elemental sulfur (S<sup>0</sup>) was 0.07 % (Table 3-3). The acid volatile sulfide (S<sub>AV</sub>) content was initially <0.01 %, and it appears, therefore, that monosulfide formation has started to occur in this sample during the tests. The sample originally contained 0.13 % S<sub>CR</sub> (Grealish *et al.* 2011) and it, therefore, appears that pyrite has not formed in the sample during the experiment (although it is possible that the original pyrite may have oxidised prior to the experiments). Dissolved sulfide was present at a very high concentration (Table 3-3), consistent with the reducing nature of the soils. The low pH (possibly due to the presence of added sucrose) may be a factor in slowing the precipitation of sulfide.



**Figure 3-5 pH and Eh dynamics during monosulfide formation potential tests in surface soil sample CNK 1.1 from Craignook wetland.**

### 3.1.4. Mineral identification by x-ray diffraction

No surface mineral efflorescences were identified or sampled at this wetland during the Phase 1 field survey.

## 3.2. Interpretation and discussion of results

The reactive metals and contaminant and metalloid dynamics tests undertaken as part of this Phase 2 assessment assists in determining the impacts on water quality by simulating the release of metal and metalloid concentrations that may occur under saturated conditions.

The 24 hour **reactive metals** studies provide an indication of those metals and metalloids which are more strongly bound to minerals (or weakly soluble with an acid extraction), and thus have the potential to be released. The use of a moderately strong acid (HCl) should provide an indication of “stored metals” and metalloids associated with iron (Fe) and manganese (Mn) oxides and organic materials as well as acid soluble minerals. It is commonly found that the concentrations of metals and metalloids released using extractions are much higher than those found in solution (Goody *et al.* 1995). Although guideline values exist for soils and sediments, these are generally for total soil concentrations, and therefore, are not directly appropriate for the data from metal mobilisation studies. Nevertheless, they provide a basis for comparison; and concentrations close to or above guideline values indicate an elevated hazard.

The metal and metalloid concentrations were all below sediment quality guidelines (SQG) and soil ecological investigation level (EIL) values (Table 3-1). The concentrations for most reactive metals and metalloids are relatively low, but relatively high for iron (Fe) (Table 3-1). Although concentrations do not exceed sediment quality guidelines (SQG) and soil ecological investigation level (EIL) values, they are generally sufficiently high ( $\text{mg kg}^{-1}$ ) to pose a potential risk to waters if dissolved, as guideline values for waters are generally of the order of  $\mu\text{g l}^{-1}$ .

The **contaminant and metalloid dynamics** was designed to determine the release of metals and metalloids in soils. The data represent the availability of metals and metalloids from a weak extraction (water, and thus easily bioavailable) of saturated soils, and for dry wetland soils (especially below Lock 1), those easily mobilised from mineral surfaces and readily soluble mineral phases (such as salts). The exercise was undertaken in a batch process for time periods of 1 day, 7 days, 14 days, and 35 days. This approach was aimed at understanding changes in concentrations over time. This is particularly important for dried soils which have been in contact with the atmosphere. The soil materials and the release/uptake of metals/metalloids are expected to change as the chemical environment changes from oxidising to reducing. Typical changes would be a reduction in redox potential (Eh), providing sufficient organic matter or other reducing agents are present, and an increase in pH (providing the soils contain or have the capacity to generate acid neutralising agents). The data can be compared to existing water quality guidelines, although care should be taken when extrapolating to surface waters without knowledge of hydrological conditions and natural chemical barriers. The impact on surface waters will be governed by the upward chemical flux which is a function of soil type, water flow, diffusion and the chemistry of the soils near the sediment-water interface. The mobility of most metals is commonly related to the stability of iron (Fe) and manganese (Mn) minerals. Under oxidising conditions iron (Fe) and manganese (Mn) oxide minerals are important sorbents for trace metals, whilst under very reducing conditions they may be incorporated into sulfide minerals. However, under moderately reducing conditions i.e. during the transition (suboxic) from oxidising to reducing conditions, iron (Fe) and manganese (Mn) are soluble and this is the period where metals may be released into solution and pose the greatest hazard.

The soil water pH was variable in the samples studied, but increased over time over the 35 day period. All samples showed a significant decrease in Eh (Figure 3-1), and for one sample, this was sufficient to allow iron (Fe) to be soluble. Iron (Fe) concentrations increased



over time in CNK 1.1, and appear to be strongly controlled by the decrease in Eh. Manganese (Mn) was also high in this sample reflecting the low Eh. Aluminium (Al) concentrations were not particularly high considering the slightly acidic nature of the soils. It was higher in the samples from the marginal sands, but not in the clay sample. Limited availability for the clays was suggested by the low concentrations in the reactive metals tests (Table 3-1). The risks associated with metal and metalloid release in Craignook wetland are probably related to the dissolution of iron and manganese compounds. The trends for most metals and metalloids are similar to iron (Fe) and manganese (Mn) (Figure 3-1 to Figure 3-3). The solubility of these elements is controlled dominantly by pH and Eh, and in the early stages of rewetting, it appears that Eh is the dominant control.

The degree to which samples exceed guideline concentrations has been used to assign a degree of hazard (Table 3-4). For some samples which required dilution, the detection limits were slightly above ANZECC/ARMCANZ environmental protection guideline values due to required dilution. Antimony (Sb), for example, was below detection limit for all samples (detection limits varying between 1 and 20  $\mu\text{g l}^{-1}$ ) as was silver, chromium, copper and lead in a few samples. It is therefore not possible to group these in Table 3-4, although it can be concluded that they either sit in the 'No hazard' or 'Low Hazard' grouping. The data are shown in Appendix 1 which displays the detection limits for individual analyses.

The data are consistent with the moderate to high net acidities noted by Grealish *et al.* (2011). The contaminant and metalloid dynamics data suggest that most soils may take some time to recover in terms of acidity. For the sandy soil CNK 2, the pH remained relatively low, but this is probably due to very limited buffering in the sand. The net acidity in soil layers of this profile were low (-65 to 20 mol  $\text{H}^+$ /tonne), and therefore recovery with added alkalinity may be relatively rapid. For the clays, the higher pH in the surface layer, high dissolved sulfide and reducing conditions, will help minimise or ameliorate the upward flux of acidity and metals from this area. Higher pH will also limit the solubilities of most trace metals. The main control on metal and metalloid mobility appears to be Eh in the Craignook wetland soils. The source of many metals appears to be closely related to iron, which may imply a source from iron oxides/oxyhydroxides. The mobility of both metals and metalloids may be a significant hazard over longer timescales, at least until reduction leads to precipitation of sulfides, in which case they may be scavenged by the precipitating iron sulfides.

**Table 3-4 Summary of the degree of hazard associated with the measured contaminant and metalloid concentrations in the Craignook wetland.**

Degree of Hazard	Guideline Threshold	Metal/Metalloid
No Hazard	Value below ANZECC/ARMCANZ guideline threshold	Se, V
Low Hazard	Value exceeds ANZECC/ARMCANZ guideline threshold, but is less than 10x exceedance	Al, As, Cd, Mn, Ni, Zn
Moderate Hazard	Value exceeds ANZECC/ARMCANZ guideline threshold by 10x or more, but is less than 100x exceedance	Co, Fe
High Hazard	Value exceeds ANZECC/ARMCANZ guideline threshold by 100x or more	

Note: Ag, Cr, Cu, Pb and Sb below detection limits in some samples due to dilutions, and therefore are likely to be in the low or no hazard classification.

The monosulfide formation potential tests assist in determining the propensity for monosulfides to form during future inundation. The sample used for this test contained 0.01% acid volatile sulfide ( $S_{AV}$ ). This provides a monosulfide formation potential hazard as 'Low hazard' (Table 3-5).

**Table 3-5 Guideline thresholds for the degree of hazard associated with acid volatile sulfide ( $S_{AV}$ ) concentrations.**

Degree of Hazard	Guideline Threshold
No Hazard	< 0.01 % $S_{AV}$
Low Hazard	0.01 % $S_{AV}$
Moderate Hazard	>0.01 – 0.05 % $S_{AV}$
High Hazard	>0.05 % $S_{AV}$

## 4. RISK ASSESSMENT

### 4.1. Risk assessment framework

Risk is a measure of both the consequences of a hazard occurring, and the likelihood of its occurrence (MDBA 2011). According to the National Environment Protection Measures (NEPM), risk is defined as "*the probability in a certain timeframe that an adverse outcome will occur in a person, a group of people, plants, animals and/or the ecology of a specified area that is exposed to a particular dose or concentration of a hazardous agent, i.e. it depends on both the level of toxicity of hazardous agent and the level of exposure*" (NEPC 1999).

The MDB Acid Sulfate Soils Risk Assessment Project developed a framework for determining risks to wetland values from acid sulfate soil hazards (MDBA 2011). The risk assessment framework has been applied in this study to determine the specific risks associated with acidification, contaminant mobilisation and de-oxygenation. In this risk assessment framework, a series of standardised tables are used to define and assess risk (MDBA 2011). The tables determine the consequence of a hazard occurring (Table 4-1), and a likelihood rating for the disturbance scenario for each hazard (Table 4-2). These two factors are then combined in a risk assessment matrix to determine the level of risk (Table 4-3).

Table 4-1 determines the level of consequence of a hazard occurring, ranging from insignificant to extreme, and primarily takes account of the environmental and water quality impacts to the wetland values and/or adjacent waters.

**Table 4-1 Standardised table used to determine the consequences of a hazard occurring, from MDBA (2011).**

Descriptor	Definition
Extreme	Irreversible damage to wetland environmental values and/or adjacent waters; localised species extinction; permanent loss of drinking water (including stock and domestic) supplies.
Major	Long-term damage to wetland environmental values and/or adjacent waters; significant impacts on listed species; significant impacts on drinking water (including stock and domestic) supplies.
Moderate	Short-term damage to wetland environmental values and/or adjacent waters; short-term impacts on species and/or drinking water (including stock and domestic) supplies.
Minor	Localised short-term damage to wetland environmental values and/or adjacent waters; temporary loss of drinking water (including stock and domestic) supplies.
Insignificant	Negligible impact on wetland environmental values and/or adjacent waters; no detectable impacts on species.

Table 4-2 determines the likelihood (i.e. probability) of disturbance for each hazard, ranging from rare to almost certain. This requires an understanding of the nature and severity of the materials (including the extent of the acid sulfate soils, the acid generating potential and the buffering capacity of the wetland soil materials) as well as contributing factors influencing the risk (MDBA 2011). Examples of disturbance include: (i) rewetting of acid sulfate soil materials

after oxidation, (ii) acid sulfate soil materials that are currently inundated and may be oxidised, or (iii) acid sulfate soil materials that are currently inundated and may be dispersed by flushing (e.g. scouring flows) (MDBA 2011). As mentioned previously, the consequence of a hazard occurring and the likelihood rating for the disturbance scenario for each hazard are then ranked using a standardised risk assessment matrix (Table 4-3).

**Table 4-2 Likelihood ratings for the disturbance scenario, from MDBA (2011).**

Descriptor	Definition
Almost certain	Disturbance is expected to occur in most circumstances
Likely	Disturbance will probably occur in most circumstances
Possible	Disturbance might occur at some time
Unlikely	Disturbance could occur at some time
Rare	Disturbance may occur only in exceptional circumstances

**Table 4-3 Risk assessment matrix, adapted from Standards Australia & Standards New Zealand (2004).**

Likelihood category	Consequences category				
	Extreme	Major	Moderate	Minor	Insignificant
Almost certain	Very High	Very High	High	Medium	Low
Likely	Very High	High	Medium	Medium	Low
Possible	High	High	Medium	Low	Low
Unlikely	High	Medium	Medium	Low	Very low
Rare	High	Medium	Low	Very low	Very low

It is suggested that:

- For very high risk immediate action is recommended.
- For high risk senior management attention is probably needed.
- Where a medium risk is identified management action may be recommended.
- Where the risk is low or very low, routine condition monitoring is suggested.

These categories of management responses have been kept quite broad to acknowledge that jurisdictional authorities and wetland managers may choose to adopt different approaches in dealing with acid sulfate soils. The imprecise nature of these management responses is intended to provide flexibility in jurisdictional and wetland manager responses to the risk ratings associated with the acid sulfate soil hazards (MDBA 2011).

## 4.2. Assessment of risks

Realisation of the main risks associated with acid sulfate soil hazards (acidification, contaminant mobilisation and deoxygenation) is highly dependent on transport and therefore on the surface and sub-surface hydrology. The risks are thus scenario dependent, and difficult to quantify without predicted changes of water flows and inputs and hydrogeological controls.

The consequences of a hazard, as outlined in Table 4-1, relate to reversible or irreversible damage to wetland values. Few studies have documented in sufficient detail the short or long term damage to inland wetland ecosystems and values caused by acid sulfate soil hazards, but short term consequences have been clearly illustrated e.g. for water quality and ecosystem impacts (McCarthy *et al.* 2006; Shand *et al.* 2010). Irreversible damage is difficult to assess due to lack of sufficient data over longer timescales and lack of knowledge, for example, on sub-surface soil recovery and metal mobilisation impacts on benthic organisms. Nevertheless, the following sections detail the hazards and likelihood of a number of scenarios and discuss consequences based on limited previous work (e.g. McCarthy *et al.* 2006; Shand *et al.* 2010). The risks to soil water quality and surface water quality are necessarily different. The risks to soil water quality in terms of acidification and contaminant release are easier to assess from the tests carried out in this study than the risks posed to surface water quality. The impacts on surface water quality will be largely controlled by upward flux of acidity and metals from the soils and sediments into the water column. This will be controlled by *inter alia* surface water volume and groundwater connectivity and level, soil type, hydraulic conductivity, degree and depth of soil cracking.

The Craignook wetland has been classified as medium conservation status by the SA Murray-Darling Basin Natural Resources Management Board (Miles *et al.* 2010).

### 4.2.1. Risks associated with acidification

The low to moderate net acidities in the dry and partially wet soils from Craignook wetland (Grealish *et al.* 2010) show that the acidification hazard is variable. Although the sandy marginal soils displayed less of an increase in pH, the net acidities are such that imported alkalinity during refilling will neutralise this acidity, especially if flows are significant. The highest hazard is probably from the clays, which had higher net acidities (-29 to 84 mol H<sup>+</sup>/tonne). However the contaminant and metalloid dynamics data suggest that the surface soils will quickly recover and become reducing, providing some protection for surface water acidification. Recovery, however, will depend largely on hydrological conditions e.g. high flows will provide some buffering and allow the acidity flux to move downwards in the soil profile. The acidification hazard is therefore considered to be moderate and probably localised in the wetland.

It is concluded that soil acidification would pose a short-term problem in the soils over much of the wetland as suggested by the measured low pH over the 35 days of the contaminant and metalloid dynamics tests. The wetland was dry during sampling, and due to its location adjacent to the river and connectivity, the likelihood of disturbance is considered **almost certain** as flows return to normal in the future. The consequences for soil ecology are likely to be short term and localised in nature and the timescale for soil recovery from acidification will probably be of the order of at least months even if there are sufficient flows, as indicated in the contaminant and metalloid dynamics experiments. A **minor** rating is therefore applied for consequence as short-term damage to soil water chemistry is considered likely. This provides a *risk rating for soil acidification* of **medium** (Table 4-4). A rating for surface water acidification will depend on surface and sub-surface hydrology. The highest risk is likely to be during low flows where the soil to water ratio is high: acidity will be most concentrated. The risk to surface water acidification is considered lowest where high flows are available to both dilute acidity and transport acidity downwards in the soil profile. Surface water acidification is likely to be lower than soil acidification, due to limited transport and buffering reactions at the soil/water interface (where recovery may be rapid), therefore an **insignificant to minor**

categorisation is given for consequence. The *risk to surface water acidification* is therefore likely to be **low to medium** (Table 4-4).

#### 4.2.2. Risks associated with contaminant mobilisation

The risks of metal and metalloid mobilisation are controlled primarily by metal abundance and availability, geochemical controls on speciation and transport mechanisms. The master variables pH and Eh exert a direct major influence on the solubility of individual metals and metalloids and minerals such as iron (Fe) and manganese (Mn) oxides and hydroxides which are important sorbents of metal and metalloid species. The moderate to high but variable acidification hazard due to the oxidation of sulfide minerals means that metals and metalloids are likely to be present in some soils, depending on the pH and Eh conditions. The data suggest that iron (Fe) and cobalt (Co) are the elements with the highest hazard, although aluminium (Al), arsenic (As), manganese (Mn), nickel (Ni) and zinc (Zn) were also relatively high (Table 3-2). The samples were variable in their responses to rewetting, largely due to differences in redox potential (Eh). Elevated concentrations of a number of metals and metalloids appear to be controlled by the reductive dissolution of an iron-bearing mineral, probably an oxide/oxyhydroxide. The risks may be higher for metalloids as both iron (Fe) and arsenic (As) were increasing in surface sample CNK 1.1 on day 35. However, further reduction processes may lead to reincorporation of metals and metalloids into sulfide minerals (following sulfate reduction), but the timescales for this are not known.

Although the timescales cannot be assessed with existing information, the data suggest that metal availability is significant for some metals. Comparisons with other studies (e.g. Nelwart Lagoon, Shand *et al.* 2010), suggest that at the pH levels noted in the contaminant and metalloid dynamics experiments for most samples, reductive processes may occur rapidly once initiated, and soil recovery may be rapid. A **moderate** rating is applied for consequence due to the presence of a number of metals and metalloids released from some of the clay soils samples, particularly as the clay soils are the dominant soil type in the wetland. Along with a likelihood of disturbance of **almost certain** as flows return to normal in the future, this provides a risk rating for contaminant mobilisation in soils of **high** (Table 4-4).

A rating for surface water impacts from metals and metalloids will depend on surface and sub-surface hydrology. The acidic to circumneutral pH values in this study, however, means that longer term impacts are possible. Chemical reactions with soils and interactions at the soil/water interface are likely to diminish any minor hazards from metal flux. The highest risk is likely to be during low flows where the soil to water ratio is high: metals will be most concentrated. The risk to surface metal and metalloid flux is considered lowest where high flows are available to both dilute metal and metalloid concentrations and transport these downwards in the soil profile. Due to enhanced mobility of metalloids at higher pH, the hazard cannot be assumed to be insignificant with the limited time series data available in this study, hence a **minor to moderate** consequence is applied. The risk to surface waters from metal mobilisation is therefore considered to be **medium to high** (Table 4-4).

#### 4.2.3. Risks associated with de-oxygenation

Monosulfidic materials are considered the main cause of deoxygenation risk in acid sulfate soils. Monosulfidic black ooze was identified in the wetland during the Phase 1 survey (Grealish *et al.* 2011).

The hazard for monosulfidic formation potential is low with acid volatile sulfide ( $S_{AV}$ ) being present with a concentration of 0.01%. Along with very high dissolved sulfide and presence of elemental sulfur ( $S_0$ ) (Table 3-3), this suggests that risks from deoxygenation could be significant. The risk of deoxygenation is most significant if the surface soils are disturbed and the soils have a consistency that will allow rapid movement and transport to areas of high value. Craignook wetland is separated from the river by a raised bank with a connection at the downstream end of the wetland. The consequence of deoxygenation is considered to be **moderate** as disturbance may lead to short term deoxygenation, particularly as the low

density surface monosulfidic soils can be mobilised easily. Taking into account the **almost certain** likelihood and the moderate consequence, a risk rating of **high** is attributed for deoxygenation potential (Table 4-4).

**Table 4-4 Summary of risks associated with acid sulfate soil materials in the Craignook wetland.**

Acidification Risk		Contaminant mobilisation		Deoxygenation
<i>Soil</i>	<i>Water</i>	<i>Soil</i>	<i>Water</i>	
Medium	Low-Medium	High	Medium-High	High

## 5. BROAD ACID SULFATE SOIL MANAGEMENT OPTIONS

The options available for rehabilitation of inland waterways containing acid sulfate soils has recently been reviewed (Baldwin & Fraser 2009) and incorporated into the *National guidance on managing acid sulfate soils in inland aquatic ecosystems* (EPHC & NRMCC 2011; see Table 5-1). The national guidance document provides a hierarchy of management options for managing acid sulfate soils in inland aquatic ecosystems including:

1. *Minimising the formation of acid sulfate soils in inland aquatic ecosystems.*
2. *Preventing oxidation of acid sulfate soils, if they are already present in quantities of concern or controlled oxidation to remove acid sulfate soils if levels are a concern but the water and soil has adequate neutralising capacity.*
3. *Controlling or treating acidification if oxidation of acid sulfate soils does occur.*
4. *Protecting connected aquatic ecosystems/other parts of the environment if treatment of the directly affected aquatic ecosystem is not feasible.*
5. *Limited further intervention.*

In designing a management strategy for dealing with acid sulfate soils in affected inland wetlands, other values and uses of a wetland need to be taken into account to ensure that any intervention is compatible with other management plans and objectives for the wetland. The medium conservation status for this wetland suggests that the management responses required should align with those suggested following the risk assessment ratings (Table 4-3).

A number of options for treating acid sulfate soils in inland wetlands have been identified (see Table 5-1). By far the best option is not to allow acid sulfate soils to build up in the first instance. This requires removing the source of sulfate from the wetland, for example, by lowering saline water tables and/or introducing frequent wetting and drying cycles to the wetland so that the amount of sulfidic material that can build up in the sediments during wet phases is limited, hence reducing the likely environmental damage (acidification, metal release or deoxygenation) that would occur as a consequence of drying.

If acid sulfate soils have formed, prevention of oxidation, usually by keeping the sediments inundated to sufficient depth, is a potential strategy. If oxidation of acid sulfate soils occurs and the sediment and/or water column acidifies, neutralisation may be necessary.

The highest risks identified in this study are due to contaminant mobilisation and deoxygenation. The likelihood of water refilling the wetland is high as flows return to normal levels. The limited number of case studies on refilling wetlands makes prediction of risk difficult in terms of determining whether reversible or irreversible damage is likely to occur. However, the low pH's observed and significant increase in pH over the 35 day period along with metal and metalloid mobilisation implies that any risks are likely to be significant.

As the wetland has previously dried and undergone oxidation, management options 1 and 2 in Table 5-1 are not relevant to the current study, although minimising further oxidation could have been an option prior to recent high flows down the River Murray. Treatment options currently remain a viable option should water quality impacts e.g. acidification of surface water and/or high metal concentrations be seen. Since the risks are scenario dependent, it is recommended that surface water monitoring be undertaken at this wetland. Based on the data from this study and elsewhere (Shand *et al.* 2010), it is likely that soil recovery will be relatively slow in the sub-surface soils as pH is relatively low in three of the four samples. It is anticipated, however, that for deeper soil layers which remain sulfuric, soil acidity may remain for periods of several months. The impacts on surface and sub-surface ecosystems are not well understood and are worthy of further work, particularly long term impacts on ecosystem functionality and diversity.



**Table 5-1 Summary of management options and possible activities, from EPHC & NRMMC (2011).**

Management Objective	Activities
<p><b>1. Minimising the formation of acid sulfate soils in inland aquatic ecosystems</b></p>	<p><b>Reduce secondary salinisation through:</b></p> <ul style="list-style-type: none"> <li>• Lowering saline water tables</li> <li>• Maintaining the freshwater lens between saline groundwater and the aquatic ecosystem</li> <li>• Stopping the delivery of irrigation return water</li> <li>• Incorporating a more natural flow regime.</li> </ul>
<p><b>2. Preventing oxidation of acid sulfate soils or controlled oxidation to remove acid sulfate soils</b></p>	<p><b>Preventing oxidation:</b></p> <ul style="list-style-type: none"> <li>• Keep the sediments covered by water</li> <li>• Avoid flow regimes that could re-suspend sediments.</li> </ul> <p><b>Controlled oxidation:</b></p> <ul style="list-style-type: none"> <li>• Assess whether neutralising capacity of the sediments and water far exceeds the acidity produced by oxidation</li> <li>• Assess the risk of deoxygenation and metal release. Monitor intervention and have a contingency plan to ensure avoidance of these risks.</li> </ul>
<p><b>3. Controlling or treating acidification</b></p>	<ul style="list-style-type: none"> <li>• Neutralise water column and/or sediments by adding chemical ameliorants</li> <li>• Add organic matter to promote bioremediation by micro-organisms</li> <li>• Use stored alkalinity in the ecosystem.</li> </ul>
<p><b>4. Protecting adjacent or downstream environments if treatment of the affected aquatic ecosystem is not feasible</b></p>	<ul style="list-style-type: none"> <li>• Isolate the site</li> <li>• Neutralise and dilute surface water</li> <li>• Treat discharge waters by neutralisation or biological treatment.</li> </ul>
<p><b>5. Limited further intervention</b></p>	<ul style="list-style-type: none"> <li>• Assess risk</li> <li>• Communicate with stakeholders</li> <li>• Undertake monitoring</li> <li>• Assess responsibilities and obligations and take action as required.</li> </ul>

## REFERENCES

- ANZECC/ARMCANZ 2000 *Australian and New Zealand Guidelines for Fresh and Marine Water Quality*, Australian and New Zealand Environment and Conservation Council and agriculture and Resource Management Council of Australia and New Zealand, Canberra.
- APHA 2005 Standard methods for the examination of water and wastewater (21st Ed.). (American Public Health Association - American Water Works Association: Baltimore, USA).
- Baldwin D and Fraser M 2009 Rehabilitation options for inland waterways impacted by sulfidic sediments. *Journal of Environmental Management* 91:311–319.
- Claff SR, Sullivan LA, Burton ED & Bush RT 2010 A sequential extraction procedure for acid sulfate soils: Partitioning of iron. *Geoderma* **155**, 224-230.
- Cline ID 1969 Spectrophotometric determination of hydrogen sulfide in natural waters. *Limnology and Oceanography* **14**, 454-458.
- EPHC & NRMCC (Environment Protection and Heritage Council and Natural Resource Management Ministerial Council) 2011 National Guidance for the management of acid sulfate soils in inland aquatic ecosystems, Canberra, ACT.
- Fitzpatrick RW, Shand P, Thomas M, Grealish G, McClure S, Merry RH & Baker A 2010 Acid Sulfate Soil investigations of vertical and lateral changes with time in five managed wetlands between Lock 1 and Wellington. CSIRO Land and Water Science Report 03/10.
- Fitzpatrick RW, Shand P, Thomas M, Merry RH, Raven MD & Simpson S 2008 Acid sulfate soils in subaqueous, waterlogged and drained soil environments of nine wetlands below Blanchetown (Lock 1), South Australia: properties, genesis, risks and management. CSIRO Land and Water Science Report 42/08.
- Goody DC, Shand P, Kinniburgh DG & Van Riemsdijk WH 1995 Field-based partition coefficients for trace elements in soil solutions. *Journal of Soil Science*, **46**, 265-285.
- Grealish G, Fitzpatrick RW & Shand P 2011 Assessment of Acid Sulfate Soil Materials in the Lock 1 to Wellington Region of the Murray-Darling Basin. CSIRO: Water for a Healthy Country National Research Flagship.
- Grealish G, Shand P, Grocke S, Baker A, Fitzpatrick R & Hicks W 2010 Assessment of Acid Sulfate Soil Materials in Lock 1 to Lock 5 Region of the Murray-Darling Basin. CSIRO: Water for a Healthy Country National Research Flagship. 769 pp.
- McCarthy B, Conalin A, D'Santos P & Baldwin DS 2006 Acidification, salinisation and fish kills at an inland wetland in south-eastern Australia following partial drying. *Ecological Management and Restoration*, vol. 7, pp. 218–23.
- McGuire MM & Hamers RJ 2000 Extraction and quantitative analysis of elemental sulfur from sulfide mineral surfaces by high-performance liquid chromatography. *Environmental Science & Technology* **34**, 4651-4655.
- MDBA 2010 Detailed Assessment of Acid Sulfate Soils in the Murray–Darling Basin: Protocols for sampling, field characterisation, laboratory analysis and data presentation. MDBA Publication No. 57/10, 58 pp.

MDBA 2011 Acid sulfate soils in the Murray–Darling Basin, Murray–Darling Basin Authority, Canberra. MDBA Publication No. 147/11.

Miles M, Wainwright P, Gonzalez D, Turner R, Frankiewicz D & Jones L 2010 Application of the South Australian River Murray wetland prioritisation methodology, Department for Environment and Heritage, Adelaide.

NEPC 1999 National environment protection (assessment of site contamination) measure 1999. National Environment Protection Council.

Shand P, Merry R, Grocke S, Thomas M, Fitzpatrick RW, Thomas B, Baker A & Creeper N 2010 Water and soil quality impacts during reflooding of Nelwart Lagoon, South Australia. CSIRO: Water for a Healthy Country National Research Flagship. 158 pp.

Standards Australia & Standards New Zealand 2004, HB 436: 2004, Risk Management, Sydney, NSW.

# APPENDICES

## APPENDIX 1 REACTIVE METALS DATA

Craignook wetland

Sample	Depth	Analysis	Ag*	Al	As	Cd*	Co	Cr*	Cu	Fe	Mn	Ni	Pb	Sb*	Se*	V	Zn
CNK 1.1	0-10	a	6.1	376	2.1	97	3.1	118	4.8	1004	255	5.7	3.7	< 2.0	58	12	8.4
		b	6.9	367	2.1	98	3.1	118	4.9	1008	255	6.1	3.8	< 2.0	58	12	8.2
CNK 1.2	10-20	a	5.9	258	1.8	36	0.51	66	3.1	781	31	3.3	2.3	2.4	42	8.1	2.9
		b	6.4	338	1.8	62	0.64	85	4.1	1185	50	4.5	2.8	< 2.4	36	13	4.5
CNK 2.1	0-5	a	0.34	43	0.59	1.0	0.21	21	0.26	77	3.0	0.38	0.02	< 1.7	2.8	0.24	0.31
		b	0.34	41	0.62	1.0	0.22	21	0.24	80	3.1	0.45	0.05	< 1.7	2.8	0.26	0.31
CNK 2.2	5-35	a	1.5	48	0.48	2.2	0.33	29	0.33	147	4.0	0.48	0.12	< 1.8	5.9	0.28	0.33
		b	0.92	52	0.45	2.2	0.36	29	0.29	128	4.1	0.46	0.18	< 1.8	2.9	0.35	0.39

Units are in mg kg<sup>-1</sup> unless indicated as below

\* Units are in µg kg<sup>-1</sup>

< value is below detection limit

## APPENDIX 2 CONTAMINANT AND METALLOID DYNAMICS DATA

Craignook wetland

Sample	Day	Depth cm	Analysis	Eh mV	EC $\mu$ S/cm	pH	Ag $\mu$ g/L	Al mg/L	As $\mu$ g/L	Cd $\mu$ g/L	Co $\mu$ g/L	Cr $\mu$ g/L	Cu $\mu$ g/L	Fe mg/L	Mn $\mu$ g/L	Ni $\mu$ g/L	Pb $\mu$ g/L	Sb $\mu$ g/L	Se $\mu$ g/L	V $\mu$ g/L	Zn $\mu$ g/L
CNK 1.1	1	0-10	a	489	2990	5.68	<0.1	<0.05	<2	0.64	13	<3	<10	<0.1	3556	<10	<6	<10	0.93	<3	12
			b	479	2541	5.54	<0.1	<0.05	<2	0.36	9.0	<3	<10	<0.1	2695	<10	<6	<10	0.74	<3	7.8
	7		a	424	3380	5.30	<0.2	<0.05	8.0	<0.5	29	<1	<3	9.0	5476	15	<3	<20	<0.6	<3	12
			b	374	3620	5.78	<0.2	<0.05	12	<0.5	29	<1	<3	10	6018	16	<3	<20	<0.6	<3	12
	14		a	234	3020	6.39	<0.02	<0.05	27	<0.3	27	<1	<3	12	5686	18	<4	<6	1.0	1.8	6.0
			b	294	3110	6.10	<0.02	<0.05	15	0.30	29	<1	<3	6.1	5734	17	<4	<6	1.0	<0.9	18
35	a	84	3180	6.55	<0.01	<0.05	46	<0.1	14	<0.9	<2	25	7010	5.6	<4	<20	0.47	3.8	<3		
	b	74	3130	6.50	<0.01	<0.05	45	<0.1	11	<0.9	<2	22	6334	6.0	<4	<20	0.39	4.3	<3		
CNK 1.2	1	10-20	a	484	933	5.74	<0.01	<0.05	0.27	0.18	0.83	<0.3	<1	<0.1	203	6.0	<0.6	<1	0.11	0.61	4.5
			b	484	372	5.54	<0.01	<0.05	0.30	0.22	2.8	<0.3	<1	<0.1	402	11	<0.6	<1	0.09	<0.3	9.1
	7		a	499	1261	4.24	<0.02	<0.05	0.20	0.06	5.4	<0.1	<0.4	<0.1	613	12	<0.4	<2	<0.08	<0.4	7.2
			b	524	1315	4.56	<0.02	<0.05	<0.2	0.12	5.9	<0.1	<0.4	<0.1	673	12	<0.4	<2	<0.08	<0.4	6.4
	14		a	364	1333	4.55	<0.01	<0.05	<0.6	0.12	7.6	<0.2	<0.6	<0.1	916	16	<0.8	<1	0.08	<0.2	11
			b	354	1073	6.63	<0.01	<0.05	0.90	0.06	0.54	<0.1	0.60	<0.1	128	4.7	<0.4	<0.6	0.10	3.7	3.6
35	a	239	590	5.01	<0.01	<0.05	1.0	0.36	15	<0.2	<0.4	0.13	834	21	<0.8	<4	0.08	<0.1	20		
	b	244	673	5.68	<0.01	<0.05	0.50	0.31	14	<0.09	0.35	0.15	996	19	<0.4	<2	0.07	<0.07	18		
CNK 2.1	1	0-5	a	534	108	4.66	<0.01	0.11	0.26	<0.03	2.3	<0.3	<1	<0.1	34	1.7	<0.6	<1	0.04	<0.3	<1
			b	539	107	4.65	<0.01	0.13	0.56	<0.03	2.2	<0.3	<1	<0.1	35	1.8	<0.6	<1	0.04	<0.3	<1
	7		a	529	79	4.56	<0.01	<0.05	0.20	<0.03	1.9	<0.07	0.20	<0.1	29	2.2	<0.2	<1	<0.04	<0.2	0.40
			b	534	83	4.53	<0.01	<0.05	0.20	<0.03	2.0	<0.07	<0.2	<0.1	30	2.4	<0.2	<1	<0.04	<0.2	0.80
	14		a	399	175	4.44	<0.01	<0.05	<0.3	<0.03	3.0	<0.1	<0.3	<0.1	48	4.0	<0.4	<0.6	0.04	0.10	1.2
			b	424	155	4.57	<0.01	<0.05	<0.3	<0.03	2.3	<0.1	<0.3	<0.1	36	3.2	<0.4	<0.6	0.04	<0.1	0.90
35	a	279	57	5.45	<0.01	0.07	<0.3	<0.01	2.9	<0.09	<0.2	<0.1	42	3.3	<0.4	<2	0.03	<0.07	1.2		
	b	289	55	5.04	<0.01	0.09	<0.3	<0.01	2.6	<0.09	<0.2	<0.1	38	3.0	<0.4	<2	0.05	0.10	1.3		
CNK 2.2	1	5-35	a	544	69	4.75	<0.01	0.10	0.35	<0.03	2.4	<0.3	1.7	<0.1	33	1.4	<0.6	<1	0.09	<0.3	<1
			b	549	64	4.77	<0.01	0.14	0.33	<0.03	2.2	<0.3	1.5	<0.1	30	1.2	<0.6	<1	0.04	<0.3	<1
	7		a	519	50	4.81	<0.01	<0.05	0.40	<0.03	2.3	<0.07	0.20	<0.1	30	2.2	<0.2	<1	<0.04	<0.2	0.40
			b	519	43	4.82	<0.01	<0.05	0.30	<0.03	2.0	<0.07	<0.2	<0.1	26	1.9	<0.2	<1	<0.04	<0.2	<0.4
	14		a	434	122	4.69	<0.01	<0.05	0.30	<0.03	3.6	<0.1	<0.3	<0.1	50	4.0	<0.4	<0.6	0.04	<0.1	1.2
			b	449	112	4.80	<0.01	<0.05	0.30	<0.03	3.0	<0.1	<0.3	<0.1	40	3.5	<0.4	<0.6	0.02	<0.1	0.60
35	a	304	37	5.31	<0.01	0.11	0.46	<0.01	2.8	<0.09	<0.2	<0.1	38	2.8	<0.4	<2	0.05	0.08	0.84		
	b	334	34	5.27	<0.01	0.17	<0.3	<0.01	2.5	<0.09	<0.2	<0.1	34	2.4	<0.4	<2	0.04	0.09	0.76		

< value is below detection limit

# APPENDIX 3 MONOSULFIDE FORMATION POTENTIAL DATA

Craignook wetland

MBO Formation Potential (MBO FP) - DAY 0

IRON DATA

DAY 0

Sample No.	org	Site Name	Site ID	Total Reactive Fe (mg/kg)				Fe(II) (mg/kg)				Eh (mV)				pH			
				Replicate 1	Replicate 2	Mean	+/-	Replicate 1	Replicate 2	Mean	+/-	Replicate 1	Replicate 2	Mean	+/-	Replicate 1	Replicate 2	Mean	+/-
22	csiro	Craignook	CNK1.1	5573	6383	<b>5978</b>	405	1258	1279	<b>1268</b>	10	473	485	<b>479</b>	6	5.49	5.47	<b>5.48</b>	0.01
32	-	Blank	-	0.3	0.1	<b>0.2</b>	0.1	<0.1	<0.1	<b>&lt;0.1</b>	<0.1	183	186	<b>185</b>	2	6.17	6.10	<b>6.14</b>	0.04

MBO Formation Potential (MBO FP) - Week 7

Sample No.	org	Site Name	Site ID	AVS (‰)				Pyrite (‰)				ES (‰)				pH				Eh (mV)				Aqueous Sulfide (µg/L)			
				Replicate 1	Replicate 2	Mean	+/-	Replicate 1	Replicate 2	Mean	+/-	Replicate 1	Replicate 2	Mean	+/-	Replicate 1	Replicate 2	Mean	+/-	Replicate 1	Replicate 2	Mean	+/-	Replicate 1	Replicate 2	Mean	+/-
22	csiro	Craignook	CNK1.1	0.02	0.01	<b>0.01</b>	<0.01	0.11	0.09	<b>0.10</b>	0.01	0.06	0.07	<b>0.07</b>	0.01	4.76	4.66	<b>4.71</b>	0.05	34	16	<b>25</b>	9	49556	50822	<b>50189</b>	633
32	csiro	Blank													5.45	5.42	<b>5.44</b>	0.02	325	338	<b>331</b>	6	< 0.1	< 0.1	<b>&lt; 0.1</b>	< 0.1	



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